



18th National and 3rd International Conference of هجدهمین همایش ملی و سومین همایش Iranian Biophysical chemistry بین المللی بیوشیمی فیزیک ایران

25-26 Des, 2024, University of Hormozgan

6-6 دی ماه ۱۴۰۳، دانشگاه هرمزگان

Cloning, expression, and characterization of a novel marine L-asparaginase from *Pseudomonas aeruginosa HR03*

Fatemeh Izadpanah Qeshmi¹, Ahmad Homaei^{1*}, Khosro Khajeh², Ehsan Kamrani³, Pedro Fernandes⁴

- 1. Department of Marine Biology, Faculty of Marine Science and Technology, University of Hormozgan, P.O. Box 3995, Bandar Abbas, Iran.
- 2. Department of Biochemistry, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran.
- 3. Fisheries Department, Faculty of Marine Science and Technology, University of Hormozgan, Bandar Abbas, Iran.
- 4. Department of Bioengineering and IBB—Institute for Bioengineering and Biosciences, Instituto Superior Técnico, Universidade de Lisboa, Av. Rovisco Pais, 1049-001 Lisbon, Portugal.

Abstract

The present study focused on the cloning, expression, and characterization of marine L-asparaginase of *Pseudomonas aeruginosa HR03* isolated from fish intestine (*L.klunzingeri*). Marine *Pseudomonas aeruginosa HR03* was used for retrieving the l-asparaginase encoding gene (HR03Asnase) of size 936 bp. The gene was successfully cloned into the pET21a vector and expressed into *Escherichia coli BL21* (DE3) for characterization of the protein. The recombinant HR03Asnase enzyme was purified by affinity chromatography using nickel affinity chromatography, and the enzymatic properties of HR03Asnase, including the effects of pH and temperature on HR03Asnase activity and kinetic parameters, were determined.

The recombinant enzyme HR03Asnase showed the highest similarity to type I bacterial L-asparaginase from *Pseudomonas aeruginosa*. The three-dimensional (3D) modeling results indicate that HR03Asnase exists as a homotetramer. Also, The Molecular weight analysis using SDS-PAGE revealed ~ 35 kDa. The HR03Asnase showed optimum pH and temperature of 8.0 and 40 °C, respectively. The maximum activity of HR03Asnase was reduced by 50% at 90 °C after 10-min incubation; though, the enzyme preserved more than 20% of its activity after 30-min incubation. This enzyme also preserved almost 50% of its activity at pH 12 after 40-min incubation. The km and Vmax of the enzyme obtained with l-asparagine as substrate were 10.904 mM and $3.44 \times 10-2$ mM/min, respectively.

The recombinant HR03Asnase of marine *P. aeruginosa* may also be explored as a potential agent in pharmaceutical and food applications. The assessment of pH and temperature stability of HR03Asnase showed that the enzyme has a wide range of activity, which is a suitable characteristic for





18th National and 3rd International Conference of هجدهمین همایش ملی و سومین همایش بین المللی بیوشیمی فیزیک ایران بیوشیمی فیزیک ایران

25-26 Des, 2024, University of Hormozgan

۶-۵ دی ماه ۱۴۰۳، دانشگاه هرمزگان

its application in different industries. Overall, the results of the present study show that marine sources are promising biological reservoirs for enzymes to be used for biotechnological purposes, and marine thermostable HR03Asnase is likely a potential candidate for its future usage in the pharmaceutical and food industries.

Key words:

Purification, Cloning, E. coli, Enzyme activity, Acrylamide

References

[1] Imada A, Igarasi S, Nakahama K, Isono M. Asparaginase and glutaminase activities of microorganisms. Microbiology, 76:85–99, 1973.

[2] Homaei A. Purifcation and biochemical properties of highly efficient alkaline phosphatase from Fenneropenaeus merguiensis brain. J Mol Catal B Enzym, 118:16–22, 2015.

[3] Gulati R, Saxena R, Gupta R. A rapid plate assay for screening 1-asparaginase producing microorganisms. Lett Appl Microbiol, 24:23–26, 1997.

[4] Lu X, Chen J, Jiao L, Zhong L, Lu Z, Zhang C, Lu F. Improvement of the activity of l-asparaginase I improvement of the catalytic activity of l-asparaginase I from Bacillus megaterium H-1 by in vitro directed evolution. J Biosci Bioeng, 128:683–689, 2019.

[5] Izadpanah Qeshmi F, Javadpour S, Malekzadeh K, Tamadoni Jahromi S, Rahimzadeh M. Persian Gulf is a bioresource of potent L-asparaginase producing bacteria: isolation & molecular diferentiating. Int J Environ Res, 8:813-818, 2014.

[6] Lee S-J, Lee Y, Park G-H, Umasuthan N, Heo S-J, De Zoysa M, Jung W-K, Lee D-W, Kim H, Kang D-H. A newly identifed glutaminase-free L-asparaginase (L-ASPG86) from the marine bacterium Mesofavibacter zeaxanthinifaciens. J Microbiol Biotechnol, 26:1115–1123, 2016.

[7] Sahu MK, Poorani E, Sivakumar K, Thangaradjou T, Kannan L. Partial purifcation and antileukemic activity of L-asparaginase enzyme of the actinomycete strain LA-29 isolated from the estuarine fsh, Mugil cephalus (Linn.). J Environ Biol, 28:645, 2007a.

[8] Qeshmi FI, Homaei A, Fernandes P, Javadpour S. Marine microbial L-asparaginase: biochemistry, molecular approaches and applications in tumor therapy and in food industry. Microbiol Res, 208:99–112, 2018.

[9] Shi R, Liu Y, Mu Q, Jiang Z, Yang S. Biochemical characterization of a novel L-asparaginase from Paenibacillus barengoltzii being suitable for acrylamide reduction in potato chips and mooncakes. Int J Biol Macromol, 96:93–99, 2017.

[10] Zuo S, Zhang T, Jiang B, Mu W. Reduction of acrylamide level through blanching with treatment by an extremely thermostable L-asparaginase during French fries processing. Extremophiles, 19:841–851, 2015.